Genetic polymorphism in DNA base excision repair gene XRCC1 among medical radiation workers

Harry Nugroho Eko Surniyantoro*, Yanti Lusiyanti, Wiwin Mailana, Devita Tetriana
Center for Technology of Safety and Radiation Metrology, National Nuclear, Energy Agency of Indonesia, Jakarta, Indonesia

ABSTRACT

X-rays repair cross-complementing group 1 (XRCC1) gene is one of the gene that plays an important role in base excision repair system (BER) and DNA repair both single and double strand breaks. Individuals with XRCC1 exon 10 (Arg399Gln) gene polymorphisms and carrying 399Gln allele variants (A allele) have a greater risk of DNA damage than their wildtype, 399Arg. The aim of this study was to examine the genotype frequencies of single nucleotide polymorphisms (SNPs) of XRCC1 exon 10 among medical radiation workers. This study involved 77 samples from several hospitals in Indonesia. Genotyping of XRCC1 exon 10 gene polymorphism was performed using PCR-RFLP. Individuals carrying A allele had lower frequency than that is carrying their wildtype of 399Arg (0.39 vs. 0.61). The results indicated that 39% of medical radiation workers had a risk of repair efficiency of DNA damage and might influence an individual's risk of cancer. Ionizing radiation induces many types of damage to DNA, requiring multiple repair pathways to restore genomics integrity. Other important genes/pathways, especially those for DNA double-strand break repair, might also play a role and should be further investigated. Furthermore, polymorphisms leading to inefficient DNA repair might also be associated with late reactions to radiotherapy.

**Keywords:**
DNA damage
XRCC1
genetic polymorphism
DNA repair
medical radiation workers

*corresponding author: eko7165@gmail.com
INTRODUCTION

The role of ionizing radiation on the human body, especially against deoxyribonucleic acid (DNA) damage is very important to be investigated, because humans cannot escape from exposure to ionizing radiation in their daily activities. During this time, the use of ionizing radiation in diagnostic and treatment devices has been widely used. This contributes to the radiation dose in the population. The most affected population group is radiation workers in hospitals that are consistently exposed to low doses of ionizing radiation.

Radiation exposure consists of two types, a high doses and low doses. High doses of radiation exposure are generally known to have an effect, including cancer induction. While exposure to low-dose radiation is still less clear. Exposure to ionizing radiation to the human body causes many adverse effects, especially in cellular DNA. This DNA damage through various means and to overcome it requires the action of DNA repair enzymes to maintain the integrity of DNA. Thus, DNA repair enzymes play an important role in maintaining genomic integrity and various DNA functional damage.  

DNA damage can be caused by endogenous and exogenous factors. The main endogenous factor is reactive oxygen species (ROS) whereas exogenous damage is caused by environmental factors such as ultraviolet rays, ionizing radiation and chemicals. DNA repair pathways consist of 3 types, namely direct reversal repair, single strand break repair and double strand break repair systems. Single strand break repair (SSBR) is the most important DNA repair system. The SSBR consists of 3 types, namely baseexcision repair (BER), nucleotide excision repair (NER) and mismatch repair (MMR). BER is a simple repair system that works in cells to repair single DNA damage caused by endogenous factors, NER is a complex repair system for repairing larger areas caused by exogenous factors, whereas the MMR plays a role for Fix base pair mismatch during the post-replication process.

BER system, plays a very important role in the human body to maintain DNA integrity, prevent cancer and DNA damage. BER is involved in the repair of oxidized bases and single strand breaks DNA after exposure by ROS, including ionizing radiation. Both oxidized brass and Single strand break, formed by ROS, are a major cause of genetic stability and cellular viability. This will lead to faster mutation rates and increase chromosomal damage. ROS are produced from endogenous and exogenous sources, including ionizing radiation. The BER pathway works by removing and replacing the damaged DNA base. This process begins with the release of DNA bases that are damaged specific glycosylase DNA enzymes, followed by sugarphosphate chain cutting, apurinic/apyrimidinic (AP) site cutting by endonuclease, DNA synthesis and ligation. This biochemical reaction involves the enzyme 8-oxoguanine DNA glycosylase 1 (OGG1), AP endonuclease 1 (APE1 or APEX1), poly (ADPribose) polymerase-1 (PARP-1), polynucleotide kinase, DNA polymerase-β.

X-rays repair cross-complementing group 1 (XRCC1) gene has a large role in DNA repair, which is as a scaffolding protein in BER, DNA repair of both SSBR and double-strand breaks, maintaining genomic stability in human cells. XRCC1 protein deficiency may decrease the ability of DNA repair and lead to increased hypersensitivity to agents that damage DNA and ionizing radiations.  

XRCC1 gene is located on chromosome 19q13.2 and consists of 17 exons encoding 633 amino acids. There are 3 SNPs that occur in XRCC1, the genetic variant (C>T) in exon 6 which converts Arg to Trp (Arg194Trp), genetic variant
(G>A) in exon 9 which converts Arg to His (Arg280His) and genetic variant (G>A) in exon 10 which converts Arg to Gln (Arg399Gln). All of SNPs can decrease DNA repair activity. This study focused on the genetic variation Arg 399 Trp in an exon 10.

Single nucleotide polymorphism (SNPs) is the most common polymorphisms in humans with a frequency≥ 1% of the population. SNPs that occur in DNA repair genes result in decreased DNA repair ability, increased mutation rate and cancer risk. SNPs that occur in the XRCC1 exon 10 gene produced 2 allele variants, namely 399 Arg and 399Gln. 399Gln alleles can be a risk factor for head and neck cancer, lung adenocarcinoma, breast cancer in African-Americans, stomachs, colon and esophagus. The aim of the study was to examine the genotype frequencies of SNPs of XRCC1 exon 10 among medical radiation workers.

**FIGURE 1.** The schematic diagram shows the regions of XRCC1 gene and shows the structure of XRCC1 with the locations of single nucleotide polymorphisms: Arg194Trp, Arg280His and Arg399Gln

**MATERIALS AND METHODS**

**Blood samples processing**

Blood samples were obtained from 77 (39 male and 38 female, mean age 37.44 ± 11.65 years) radiation workers at several hospitals in Indonesia. Genomic DNA was purified from lymphocytes extracted from whole blood using the QIAamp DNA Kit (Qiagen) according to the manufacturer’s instructions. The obtained DNA was stored at -20°C until analysis. An informed consent was obtained from each subject at the start of this study.

**PCR-RFLP genotyping assays**

Genotyping of XRCC1 exon 10 gene polymorphism was performed using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP), as described previously by Andreassi et al. with the forward primer 5'- AGTAGTCTGCTGGCTCTGG-3' and the reverse primer 5'- TCTCCCTTGGTCTCACCCT-3'. The PCR reactions were carried out with denaturation of 95°C for 3 min, followed by 30 cycles of 15 sec at 95°C, 15 sec at 60°C and 15 sec at 72°C and final 1 min at 72°C. Following amplification, PCR products
were digested using 10 U of restriction enzyme \textit{MspI} (BioLabs, Inc.) for 16 hrs at 37°C, and electrophoresed on a 3% agarose gel. The wild type GG genotype for codon 399 was determined by the presence of two bands at 269 and 133 bp, the mutant heterozygous GA genotype was determined by the presence of three bands at 402, 269 and 133 bp, while the mutant homozygous AA genotype was determined by the presence of the uncut 402 bp band (indicative of the absence of the \textit{MspI} cutting site).

**Statistical Analysis**

The statistical analysis of the data were conducted with SPSS version 16.0 for windows. Data were expressed as mean ±SD (standard deviation). Characteristics of the subjects were analyzed by the Mann-Whitney U test. Genotype and allele frequencies were shown on frequencies distribution table. A \( p \) value < 0.05 was considered to be significant.

**RESULTS**

In this study, a SNPs of the \textit{XRCC1} gene Arg399Gln was investigated. The genotype analysis of these SNPs of the \textit{XRCC1} gene, for medical radiation workers from several hospitals in Indonesia was performed using PCR-RFLP method. The characteristic of subjects according to age and gender data was displayed at TABLE 1.

<table>
<thead>
<tr>
<th>Character</th>
<th>Subjects</th>
<th>( p )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td></td>
<td></td>
</tr>
<tr>
<td>• Male</td>
<td>39</td>
<td>0.909</td>
</tr>
<tr>
<td>• Female</td>
<td>38</td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>37.44 ± 11.65</td>
<td>0.527</td>
</tr>
</tbody>
</table>

This study was used 77 samples which consisted of 39 male and 38 female. Statistically, the characteristics of age and gender did not show any differences among the subjects.

Based on FIGURE 1, GG genotype (wild type) was shown with 269+133 bp fragment length, GA genotype with 402+269+133bp, AA genotype with 402bp. Overall, the variant allele frequencies were 0.61 for \textit{XRCC1} 399Arg (G allele) and 0.39 for \textit{XRCC1} 399Gln (A allele). The genotype frequencies of \textit{XRCC1} gene polymorphism showed in TABLE 2 bellow.
TABLE 2. Genotype frequencies of XRCC1 gene polymorphism (n = 77)

<table>
<thead>
<tr>
<th>Genotype</th>
<th>n</th>
<th>Frequency (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Genotype codominant</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>• GG</td>
<td>31</td>
<td>40.26</td>
</tr>
<tr>
<td>• GA</td>
<td>32</td>
<td>41.56</td>
</tr>
<tr>
<td>• AA</td>
<td>14</td>
<td>18.18</td>
</tr>
<tr>
<td><strong>Dominant</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>• GG</td>
<td>31</td>
<td>40.26</td>
</tr>
<tr>
<td>• GA+AA</td>
<td>46</td>
<td>59.74</td>
</tr>
<tr>
<td><strong>Recessive</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>• GG+GA</td>
<td>63</td>
<td>81.82</td>
</tr>
<tr>
<td>• AA</td>
<td>14</td>
<td>18.18</td>
</tr>
<tr>
<td><strong>Allele</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>• G</td>
<td>94</td>
<td>61.04</td>
</tr>
<tr>
<td>• A</td>
<td>60</td>
<td>38.96</td>
</tr>
</tbody>
</table>

**DISCUSSION**

All the genotypes distributions were in Hardy-Weinberg equilibrium. Several reports indicate that the variant alleles of the repair polymorphisms examined may truly affect DNA repair function. The SNPs in the XRCC1 exon 10 gene resulted in the substitution of a base nitrogen number 28152, guanine (G) to adenine (A) at codon number 399, resulting in amino acid conversion from arginine (CGG) into glutamine (CAG). This polymorphism is in the BRCT domain which is very important to bind PARP-1. PARP-1 will quickly bind the SSBs and will be activated soon. Activation of PARP-1 will increase DNA protein accumulation in damaged DNA. This polymorphism given will decrease the interaction of PARP-1, so DNA repair proteins can not work properly.21

Several previous studies have shown different results. Norjmaa et al.22 reported that patients with Arg/Gln (mutant heterozygous) and Gln/Gln (mutant homozygous) genotypes are at greater risk of myelodysplastic syndrome (MDS) than patients with Arg/Arg (normal) genotypes. Arg399Gln genotype carrier variants have higher levels of DNA adducts,23 greater risk of ionizing sensitivity and DNA damage associated with smoking habits.24-26 A meta-analysis study in the Asian population shows that patients with 399Gln alleles have a higher risk of breast cancer (OR = 1.54; 95% CI: 1.18-2.01).27

The radiation workers in this study, such as radiographers, X-ray machine operators and radiologists worked in installations using low-dose ionizing radiation sources. These results in susceptibility to DNA damage due to exposure to ionizing radiation, especially DNA repair gene. Ionizing radiation is the one that has high energy and is able to release ions that can break covalent bonds, which in turn can damage the structure of human DNA. Another result is the emergence of ROS that can oxidize proteins and lipids, and induce the formation of SSBs. All such damage can cause cell death.28 Ionizing radiation includes X-rays, gamma rays, alpha, beta, and neutron particles. The radiation dose is measured by the amount of radiation absorbed by 1 kg of tissue in Gray (Gy).29

The nuclear energy regulatory agency of Indonesia arranges the average effective dose limit value for radiation workers is 20 mSv per year averaged over 5 years. It means that doses accumulated for 5 years should not exceed 100 mSv. The average equivalent dose for eye pieces is 20 mSv per year, the average equivalent dose for the skin is 500 mSv per year and the equivalent dose for the hand or foot is 500 mSv per year.30

**CONCLUSION**

Individuals carrying a XRCC1 399Gln (A allele) have a lower frequency than 399Arg (0.39 vs. 0.61). Our results suggested that 39% of medical radiation workers have a risk of repair efficiency of DNA damage and might influence
an individual’s risk of cancer. Ionizing radiation induces many types of damage to DNA, requiring multiple repair pathways to restore genomics integrity. Other important genes/pathways, especially those for DNA double-strand break repair, might also play a role and should be further investigated. Furthermore, polymorphisms leading to inefficient DNA repair might also be associated with late reactions to radiotherapy.

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